$\begin{array}{c} O2^{i} - Re - O1 \\ O2^{i} - Re - O4^{i} \end{array}$	179.56 (14) 89.5 (2)	O4 ⁱ —Re—C11 O3—Re—C11	91.50 (12) 88.95 (12)			
$O1$ —Re— $O4^{1}$	90.4 (2)					
Symmetry code: (i) $-x$, $1 - y$, $-z$.						

The title structure was solved via direct methods and all non-H atoms were refined with anisotropic displacement parameters. A molecule of dichloromethane resides on a general position. H atoms were placed in idealized positions with C—H distances of 0.93 Å. The common orientation of the displacement ellipsoids of the core atoms of the molecule suggests the presence of systematic error, most probably associated with the effects of residual effects of uncorrected absorption. It is therefore to be anticipated that a small systematic error is associated with the atomic positions and necessarily with the bond lengths and angles involving these atoms.

Data collection: MSC/AFC Diffractometer Control Software (Molecular Structure Corporation, 1989). Cell refinement: MSC/AFC Diffractometer Control Software. Data reduction: TEXSAN (Molecular Structure Corporation, 1985). Program(s) used to solve structure: SHELXS86 (Sheldrick, 1990). Program(s) used to refine structure: SHELXL93 (Sheldrick, 1993). Molecular graphics: XP (Siemens, 1994). Software used to prepare material for publication: SHELXL93.

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Lists of atomic coordinates, displacement parameters, structure factors and complete geometry have been deposited with the IUCr (Reference: BS1024). Copies may be obtained through The Managing Editor, International Union of Crystallography, 5 Abbey Square, Chester CH1 2HU, England.

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6-L-Alanineferrirubin, a Ferrichrome-type Siderophore from the Fungus Aspergillus ochraceous

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Abstract

The molecular structure of 6-L-alanineferrirubin tetradecahydrate, $[Fe(C_{41}H_{64}N_9O_{16})].14H_2O$, has been determined in order to confirm its chemical structure. The structural results show that the presence of an alanine in place of a serine or a glycine at position 6 in the cyclic hexapeptide has very little effect on the conformation of the 18-membered ring or on the geometry of the octahedral iron coordination.

Comment

The ferrichrome-type siderophores are a group of ironchelating cyclic hexapeptides produced by a number of fungal species in response to low iron availability (Neilands, 1981). All the hexapeptides in this family have the general formula cyclo{Orn1-Orn2-Orn3-A-B-Gly}, where Orn1, Orn2 and Orn3 are N-acylated-N-hydroxyornithine residues, and A and B are either glycine or serine, and sometimes alanine, as in ferrichrome C (Tademura & Sato, 1967) and malinochrome (Emery, 1980). The N-acyl groups are derivatives of carboxylic acids.

Asperchromes are a group of new siderophores belonging to the ferrichrome family isolated from the fungus *Aspergillus ochraceous*. Most of them are characterized by having heterogeneous *N*-acyl groups, a feature which was not observed previously in ferrichromes. The isolation and description of their chemical structures



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Fig. 1. Perspective drawing of the title molecule showing the atom-numbering scheme. The displacement ellipsoids correspond to 50% probability contours and H atoms have been omitted for clarity.

have been reported earlier (Jalal *et al.*, 1984). The crystal structures of two of the asperchromes (B1 and D1) have also been reported (Jalal, Hossain, van der Helm & Barnes, 1988). 6-L-Alanineferrirubin (asperchrome A) is different from the other asperchromes in that it has three homogeneous *N*-acyl groups, derived from *trans*-anhydromevalonic acid, as in ferrirubin (Barnes *et al.*, 1985), but differing from this compound in having the serine in position 6 replaced by alanine.

In the solid state, 6-L-alanineferrirubin displays all the common features of ferrichrome-type siderophores, the A-cis coordination for the iron and a hexapeptide ring conformation with two β -turns, *i.e.* a β (II) turn resulting in an N1-H-··O4 hydrogen bond [3.021 (7) Å] and a β (I) turn without any N4—H···O1 [4.280(8) Å] hydrogen bond. It also has the commonly observed intramolecular hydrogen bond [N2-H···O21 2.790(7)Å] linking the peptide backbone with the iron coordination. The substitution of an alanine in place of a serine at residue 6 seems to have very little effect on the conformation of the 18-membered peptide ring. The r.m.s. difference of the 18 endocyclic torsion angles of 6-L-alanineferrirubin and ferrirubin is less than 3° (Barnes et al., 1985). The iron coordination geometry in 6-L-alanineferrirubin also shows a remarkable similarity with that of the ferrirubin molecule.

Like most of the ferrichrome-type siderophores, the crystal structure of 6-L-alanineferrirubin is heavily hydrated. All the 14 water molecules are extensively hydrogen bonded among themselves and with the 6-L-alanineferrirubin molecules. There are no intermolecular hydrogen bonds.



Fig. 2. A stereoview of the packing down the crystallographic b axis.

Experimental

6-L-Alanineferrirubin was crystallized from aqueous solution by slow evaporation. The small prismatic crystals were unstable at room temperature but were found to be suitable for data collection at low temperature.

Crystal data

$[Fe(C_{41}H_{64}N_9O_{16})].14H_2O$	Mo $K\alpha$ radiation
$M_r = 1247.07$	$\lambda = 0.71073 \text{ Å}$
Monoclinic	Cell parameters from 44
P21	reflections
<i>a</i> = 19.903 (6) Å	$\theta = 10 - 14^{\circ}$
b = 13.757(5) Å	$\mu = 0.345 \text{ mm}^{-1}$
c = 10.966 (4) Å	T = 163 (2) K
$\beta = 90.39 (2)^{\circ}$	Prism
$V = 3002.5 (18) \text{ Å}^3$	$0.30 \times 0.22 \times 0.17 \text{ mm}$
Z = 2	Red
$D_x = 1.379 \text{ Mg m}^{-3}$	
D_m not measured	
Data collection	

Enraf–Nonius CAD-4	$\theta_{\rm max} = 26.44^{\circ}$			
diffractometer	$h = -24 \rightarrow 24$			
$\theta/2\theta$ scans	$k = 0 \rightarrow 17$			
Absorption correction: none	$l = 0 \rightarrow 13$			

$[Fe(C_{41}H_{64}N_9O_{16})].14H_2O$

6423 measured refl	lections	3 standard reflection	ons	O11-N11-C105	115.3 (5)	N5C501C503	109.9 (6)
6423 independent i	reflections	frequency: 120	min	012C107N11	119.1 (6)	C502C501C503	107.2 (5)
5471 reflections wi	ith	intensity decay:	4.5%	N11-C107-C108	122.0 (6)	O5C502N6	121.4 (6)
$I > 2\sigma(I)$				C102—N2—C201	118.7 (5)	N6-C502-C501	116.0 (6)
				N2-C201-C202	110.8 (5)	C502—N6—C601	122.0 (6)
Refinement				N2	112.0 (5) 109.3 (5)	N6C601C602 O6C602N1	115.9 (5) 124.0 (6)
Refinement on F^2		$(\Delta/\sigma)_{\rm max} = 0.075$	_	O2C202N3	122.2 (6)	O6-C602-C601	119.3 (6)
R(F) = 0.0648		$\Delta \rho_{\rm max} = 0.72 \ {\rm e} \ {\rm \AA}$	-3	O2-C202-C201	121.6 (6)	NIC602C601	116.7 (6)
$wR(F^2) = 0.1628$		$\Delta \rho_{\rm min}$ = -0.62 e	$Å^{-3}$	C602N1	C101C102	- 149.7 (6	ć)
S = 1.079		Extinction correct	ion: none	N1 - C101 - C102 - N2 - 167.7 (5)			
6416 reflections		Scattering factors	from	$\begin{array}{cccccccccccccccccccccccccccccccccccc$)) 7)
730 parameters		International To	ubles for				7)
H atoms: see below Crystallography (Vol.		v (Vol. C)	C201—C2	02—N3—C301	- 164.6 (5	5)	
$w = 1/[\sigma^2(F_o^2) + (0.0864P)^2]$		Absolute configuration:		C202N3	C301C302	-103.6 (8	3)
+ 6.0425P]	2	Flack (1983)		N3	$-C_{302}$ N4 $-C_{401}$	2.6 (9	7) 6)
where $P = (F_o^2)$	$+ 2F_c^2)/3$	Flack parameter =	0.02 (3)	C302—N4	-C401-C402	- 161.6 (6	5)
				N4-C401	C402N5	179.2 (6	5)
Table 1 Sal	ected apon	etric narameters (Å °)	C401C4	02—N5C501	- 175.0 (6	5)
	ecieu geom	ien ic purumeters (л,)	C402—N5 N5—C501		- 55.3 (8	3) 6)
Fe1031	1.974 (4)	C202—N3	1.349 (9)	C501	02—N6—C601	133.2 (0	5) 6)
$\frac{1}{1} = 021$	2 010 (5)	N21-C207	1.481 (9)	C502—N6	C601C602	81.2 (8	8)
Fe1-011	2.012 (4)	N3-C301	1.462 (8)	C101N1	C602C601	179.6 (6	5)
Fe1012	2.026 (4)	C301-C302	1.524 (9)	N6—C601	C602N1	3.5 (1	10)
Fe1-032	2.031 (5)	C301—C303	1.543 (9)	All the H atoms	were placed	in calculated positi	ons, riding
011—N11 012 C107	1.380 (7)	C302-03	1.241 (8)	on their attached a	toms, with a	tomic displacement of	coefficients
O21—N21	1.368 (7)	C305—N31	1.468 (7)	1.2 times the U_{eq}	values of th	ne attached atom. N	lo attempts
O22C207	1.296 (9)	N31-C307	1.323 (9)	were made to loca	te the H ator	ms of the water mol	lecules.
O31—N31	1.377 (7)	N4-C401	1.460 (8)	Data collection	: CAD-4 So	ftware (Enraf-Noni	ius, 1989).
O32—C307	1.282 (8)	C401C403	1.511 (10)	Cell refinement: lo	cal program.	Data reduction: loca	al program.
NI	1.307 (8)	C401 - C402 C402 - O4	1.331 (10)	Program(s) used	to solve stru	acture: SHELXS86	(Sheldrick,
C101—C103	1.538 (9)	C402—N5	1.330 (9)	1990). Program(s) used to	refine structure:	SHELXL93
C101-C102	1.543 (8)	C403	1.409 (9)	(Sheldrick, 1993)	. Molecula	r graphics: SHEL	XL93 and
C102—O1	1.233 (8)	N5-C501	1.481 (9)	PLUTO (Motherw	ell & Clegg,	1978).	
C102—N2 C105—N11	1.332 (8)	C501-C502	1.504 (9)				
N11C107	1.316 (9)	C502—O5	1.249 (9)	This work is s	upported by	v NIH grant No. C	GM21822-
N2—C201	1.476 (8)	C502—N6	1.323 (10)	(DvdH)	······································	, 8	
C201—C202	1.534 (9)	N6C601	1.464 (9)	(D (ull):			
$C_{201} = C_{203}$	1.529 (9)	C601 - C602 C602 - C602	1.517(10)	Lists of atomic coord	linates, displac	cement parameters, stru	ucture factors
021 Est 021	01.2 (2)	N2 C202 C201	116 1 (6)	and complete geome	try have been o	deposited with the IUC	r (Reference:
O_{31} Fe1 O_{21}	91.3 (2)	$C_{207} - N_{21} - O_{21}$	116.1 (6)	BS1004). Copies m	ay be obtaine	ed through The Mana	iging Editor,
O21—Fe1—O22	78.0 (2)	C207—N21—C205	129.5 (6)	International Union	of Crystallogra	phy. 5 Abbey Square,	Chester CH1
O31—Fe1—O11	92.0 (2)	O21-N21-C205	113.8 (5)	2HU, England.			
O21-Fe1-O11	87.2 (2)	O22C207N21	116.7 (6)				
022 - Fe1 - 011 031 - Fe1 - 012	162.7(2) 163.9(2)	N21	122.4 (6)	Deferences			
O21—Fe1—O12	101.2 (2)	C202—N3—C301	122.4 (6)	References			
O22—Fe1—O12	95.4 (2)	N3-C301-C302	113.7 (5)	Barnes, C. L., Hoss	ain, M. B., Ja	lal, M. A. F., Eng-Wi	ilmot, D. L.,
O11—Fe1—O12	78.7 (2)	N3C301C303	108.5 (5)	der Helm D (10)	Clison, D. A., P	4 CA1 341 347	ha, K. & van
031 - Fe1 - 032 021 - Fe1 - 032	78.2 (2) 169 0 (2)	$C_{302} - C_{301} - C_{303}$	112.1 (0)	Emery T (1980) R	iochem Rionh	1. C41, 541-547.	n
022—Fe1—032	99.6 (2)	O3-C302-C301	119.0 (6)	Enraf-Nonius (1989	CAD-4 Sof	tware Version 5.0 E	nraf-Nonius
O11—Fe1—O32	96.6 (2)	N4-C302-C301	117.8 (6)	Delft, The Nether	lands.		
O12—Fe1—O32	89.7 (2)	C307—N31—O31	116.1 (5)	Flack, H. D. (1983).	Acta Cryst. A	39 , 876–881.	
NII-OII-Fel C107-012-Fel	110.2 (3)	C307N31C305	129.6 (6)	Jalal, M. A. F., Hos	sain, M. B., v	/an der Helm, D. & P	Barnes, C. L.
N21-021-Fe1	111.7 (4)	O32-C307-N31	117.7 (6)	(1988). Biol. Met.	1, 77–89.		
C207-022-Fe1	114.0 (4)	O32-C307-C308	123.2 (6)	Jalal, M. A. F., Moc	herla, R., Bar	nes, C. L., Hossain, M	. B., Powell,
N31-031-Fe1	111.7 (3)	N31-C307-C308	119.0 (5)	D. R., Eng-Wilmo	ot, D. L., Gra	yson, S. L., Benson, I	B. A. & van
C307	112.8 (4)	C302—N4—C401	123.7 (6)	der Helm, D. (198	s4). J. Bacteri	<i>91.</i> 158 , 683–688.	ю р.
N1-C101-C103	112.6 (5)	N4-C401-C403	106.0 (6)	for Plotting Ma	5. & Clegg	z, w. (1978). PLUT Crustal Structures II	o. Program
N1-C101-C102	107.2 (5)	C403-C401-C402	112.5 (6)	Cambridge Engla	nd	crystal structures. U	inversity of
C103—C101—C102	113.8 (5)	O4C402N5	122.2 (6)	Neilands, J. B. (198	1). Annu. Rev.	Biochem, 50, 715-73	1.
01-C102-N2	122.0 (6)	04—C402—C401 N5—C402—C401	120.8 (6)	Sheldrick, G. M. (19	990). Acta Crv	st. A46, 467-473.	-
N2-C102-C101	120.2 (3)	042—C403—C401	113.1 (6)	Sheldrick, G. M. (19	93). SHELXL	.93. Program for the F	Refinement of
C107—N11—O11	116.9 (5)	C402N5C501	119.6 (6)	Crystal Structures	. University of	f Göttingen, Germany.	-
C107—N11—C105	127.3 (5)	N5-C501-C502	110.8 (5)	Tademura, M. & Sat	o, S. (1967).	Agric. Biol. Chem. 31,	1482–1485.